

SHORT COMMUNICATION

THE BIOSYNTHESIS OF STEROLS IN PUMPKIN

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Abstract—Tritium labeled 3β -hydroxy- 5α -stigmasta-7,24(28)Z-diene (I) was administered to pumpkin plants (*Cucurbita pepo* L.) and the sterols were isolated. The specific incorporation rates were: 3β -hydroxy- 5α -stigmasta-7,22t,25-triene (II) $6.7 \times 10^{-1}\%$, 3β -hydroxy- 5α -stigmasta-7,25-diene (III) $4.1 \times 10^{-2}\%$, and α -spinasterol (IV) $3.4 \times 10^{-4}\%$. From this result, I is tentatively regarded as a precursor for II and III.

INTRODUCTION

PUMPKIN seeds (*Cucurbita pepo* L.) contain several Δ^7 -sterols,¹ among them a 24-ethylidene compound 3β -hydroxy- 5α -stigmasta-7,24(28)Z-diene,* recently synthesized in this laboratory.² 24-Ethylidene sterols can act as key intermediates in phytosterol biogenesis.^{3,4}

RESULT AND DISCUSSION

The following radioactive stigmastan derivatives were isolated from pumpkin plants after administration of $[23,25\text{-}^3\text{H}_2]$ - 3β -hydroxy- 5α -stigmasta-7,24(28)Z-diene (I):

TABLE 1. INCORPORATION OF $[23,25\text{-}^3\text{H}_2]$ - 3β -HYDROXY- 5α -STIGMASTA-7,24(28)-DIENE INTO THE STEROLS OF PUMPKIN

Compound†	Specific incorporation rate [§] (%)
3β -Hydroxy- 5α -stigmasta-7,22t,25-triene (II)	6.7×10^{-1}
3β -Hydroxy- 5α -stigmasta-7,25-diene (III)	4.1×10^{-2}
3β -Hydroxy- 5α -stigmasta-7,22t-diene (IV) (α -spinasterol)	3.4×10^{-4}

† The configuration at C-24 is assumed to be α_F ; see, however, W. SUCROW and B. GIRGENSOHN, *Chem. Ber.* in press.

* For definition of the symbols Z and E for configuration of double bonds see J. E. BLACKWOOD, C. L. GLADYS, K. L. LOENING, A. E. PETRARCA and J. E. RUSH, *J. Am. Chem. Soc.* **90**, 509 (1968).

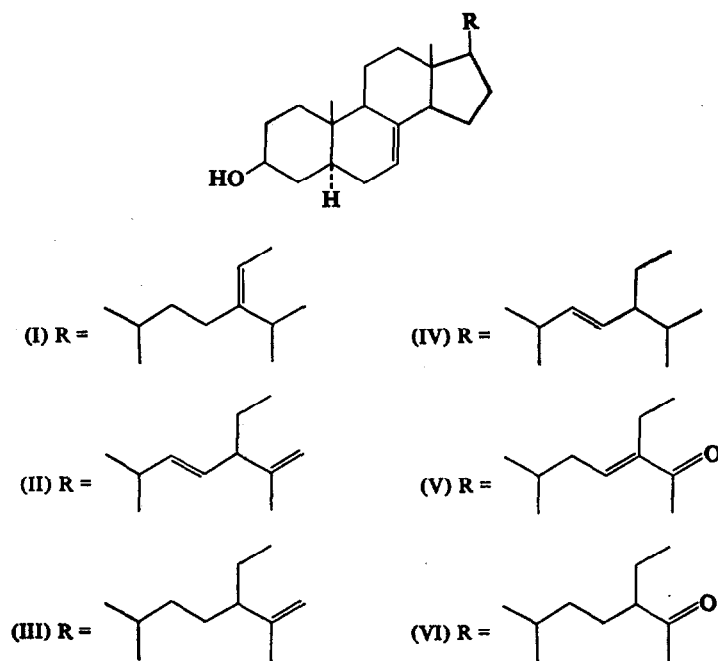
¹ W. SUCROW and A. REIMERDES, *Z. Naturforsch.* **23b**, 42 (1968).

² W. SUCROW and B. RADÜCHEL, *Chem. Ber.* **102**, 2629 (1969).

³ A. R. H. SMITH, L. J. GOAD and T. W. GOODWIN, *Biochem. J.* **104**, 56 c (1967).

⁴ D. J. BAISTED, *Phytochem.* **8**, 1697 (1969).

⁵ H. R. SCHÜTTE, *Radioaktive Isotope in der Organischen Chemie und Biochemie*, p. 86, Verlag Chemie, Weinheim (1966).



This result indicates that compounds II and III are synthesized in the plant from I as a precursor. Goodwin *et al.*³ have proposed a C-24 carbonium ion as a common intermediate in the biosynthesis of both Δ^{25} and $\Delta^{24(28)}$ -compounds. Our result shows that this mechanism should be modified for pumpkin plants since the Δ^{25} compound (II) is generated from the $\Delta^{24(28)}$ compound (I). It is, however, possible that I is converted into a C-24 carbonium ion by addition of a proton. Hydride transfer from C-25 to C-24 would yield a C-25 carbonium ion which, by proton loss from C-26, would lead to a Δ^{25} double bond. Whether II and III are formed successively from the precursor I ($\text{I} \rightarrow \text{II} \rightarrow \text{III}$) or independently ($\text{III} \leftarrow \text{I} \rightarrow \text{II}$) will be the subject of further studies by administration of labeled II and III. In our opinion, α -spinasterol (IV) is not generated directly from I because of its very small incorporation rate. Ellouz and Lenfant⁶ have administered labeled stigmastanol to *Dictyostelium discoideum* and shown that it is a precursor of 3β -hydroxy- 5α -stigmast-22-ene. According to this scheme, α -spinasterol could be biosynthesized from 3β -hydroxy- 5α -stigmast-7-ene which, however, has not yet been isolated from pumpkin.

EXPERIMENTAL

Plant Material

Pumpkin seeds were germinated for 4 days and then cultivated on Knop's solution for 12 days in a greenhouse at 22°.

Preparation of the Radioactive Precursor

3β -Acetoxy- 5α -cholest-7-ene-24-one² was treated with T_2O in basic medium.⁷ Wittig reaction with triphenyl ethylidene phosphorane and saponification yielded $[23,25\text{-}^3\text{H}_3]$ - 3β -hydroxy- 5α -stigmasta-7,24(28)Z-diene (I), specific activity 3.50×10^9 counts/min/mole.

⁶ R. ELLOUZ and M. LENFANT, *Tetrahedron Letters* 2655 (1969).

⁷ M. LINDBERG, F. GAUTSCHI and K. BLOCH, *J. Biol. Chem.* **238**, 1661 (1963).

Administration of the Labeled Compound

Twenty mg labeled I (total activity 1.70×10^8 counts/min) were emulsified in the system cotton seed oil-sucrose monostearate-water. The roots were separated from the plants and the stalks dipped into the emulsion which was consumed within 24 hr. After an additional 3 days on Knop's solution the sterol mixture was extracted by the usual procedure. Chromatography on alumina and silver nitrate impregnated silica gel⁸ yielded the sterols II-IV which were recrystallized from methanol to constant radioactivity. Because of the relatively high specific radioactivities of II and III* (3-610, 3-211, 3-100, 3-082, 3-08⁺, 3-081 $\times 10^4$ counts/min/mg for II and 7-850, 7-820, 7-819 $\times 10^3$ counts/min/mg for III) it was supposed that they still contained some radioactive precursor (I), which cannot be removed by crystallization. Therefore, II and III were converted to the ketones V and VI,⁸ which were diluted with inactive carriers (V, 1:2; VI, 1:5-5), equilibrated in basic medium and purified by preparative TLC with petrol ether-ether (6:4) on silica gel. As expected, this resulted in a loss of radioactivity. Several crystallizations from methanol yielded pure V (821, 935, 935, 928 counts/min/mg) and VI (680, 625, 630, 632 counts/min/mg). No radioactive impurities were detected by TLC in CHCl_3 -EtOAc (9:1). The specific incorporation rates were calculated to $6.1 \times 10^{-1}\%$ for II and $4.1 \times 10^{-2}\%$ for III.

Ozonization of α -spinasterol (IV) which showed only small radioactivity afforded 2-ethyl-3-methyl-butyraldehyde isolated as 2,4-dinitrophenylhydrazone⁹ and crystallized twice from methanol (42 and 40 counts/min/mg). The steroidal aldehyde 3 β -hydroxy-5 α -pregn-7-ene-21-al was isolated likewise and showed no radioactivity. Specific incorporation rate for IV is $3.4 \times 10^{-4}\%$.

Radioactivity measurements were carried out in a Beckman scintillation counter, using toluene as solvent with the usual scintillators.

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* II has been diluted 1 : 28 with inactive carrier.

⁸ W. SUCROW, *Chem. Ber.* **99**, 3559 (1966).

⁹ M. DEVYS, A. ALCAIDE, M. BARBIER and E. LEDERER, *Phytochem.* **7**, 613 (1968).